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A Spatial Model for Tumor Drug Resistance: the case of GIST Liver Metastases

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WHAT IS THE GIST ?

GIST : Gastro-Intestinal Stromal Tumors

■ In 50% of cases, the GIST are metastatic.

- In 50% of metastatic case, metastases are present in the liver.
- \Rightarrow 25% of patient with a GIST have a liver metastases.

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CURRENT TREATMENT PROTOCOL

- First line: Imatinib, a cytotoxic drug. Inhibits a specific receptor tyrosine kinase (BCR-Abl).
 - 10-15% of patients present a mutation of gene KIT (on exons [4, 5]) that immediately leads to an imatinib insensitivity (see Fig 1b).

For the other patients, the imatinib controls metastatic lesions during a period more or less long: around 20-24 months in 85% of cases (see Fig 1a).

Second-line: sunitinib, which is a multi-targeted receptor tyrosine kinase inhibitor (that inhibits PDGFRs, VEGFRs and KIT), which has both cytotoxic and antiangiogenic effects.



(A) Patient A: profile that has a good (B) Patient E: typical profile of answer on treatment - x-axis: time in resistance to Imatinib associated to a days; y-axis: the tumor area in mm².

particular genetic mutation (EXON11) - x-axis: time in days; y-axis: the tumor area in mm²

FIGURE: Evolution of a GIST for two differents patients: one representing the most common case and the other one a patient with a mutation

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THERAPEUTIC FOLLOW-UPS OF PATIENT

CT-scans (usually every 2 months)

 \rightsquigarrow Track the disease evolution and the response to the treatment.

Clinicians challenge: optimizing cancer treatments and particularly the switch time from the first-line to the second-line treatment, in order to increase the overall survival time.

RECIST criteria: the diameter of the largest lesions is the *only* information extracted from the CT-scan.

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Our aim

Int

Develop a mathematical model based on medical images of liver metastases of locally advanced GIST in order to determine, for *each* patient:

- the treatment response
- the relapse times after the first-line and the second-line treatments
- geometric specificities of tumor growth

Our model enable to:

- Compare the model with the medical images
- Highlight more crucial data such as tumor heterogeneities and geometrical properties of the tumor growth, which may provide more precise indicators than the RECIST criteria.

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The existing models

Several theoretical tumor growth models have been developed:

- ODEs model as Mendelsohn, logistic, Gompertz or Bertalanffy ([9] and in [− ref Papier Seb −]) → This kind of models fit data only on the tumor volume
- Cellular automata [1, 8]
- Models based on mixture theory [2]
- Agent-based models [10]
- Models based on fluid mechanics [7]
- Models based on reaction-diffusion theory [11]
- \Rightarrow Several scale, from the cells to the tissues, are covered by this large variety of models.

Our choice: a model based on fluid mechanics.

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- 5 DISCUSSION

CT-SCANS I

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Cross-section selected is the same for all exams.



(A) Sept 16, 2008 - Day 119

(B) June 30, 2009 - Day 406

(c) July 5, 2010 - Day 776

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CT-SCANS II



(D) Oct 25, 2010 – Day 888

(E) Jan 7, 2011 - Day 962

(F) June 10, 2011 - Day 1116

$\ensuremath{\mathbf{Figure:}}$ Spatial evolution of the patient A metastasis on a series of CT-scans

GIST are characterized by the presence of some *heterogeneity* in the tumor.

The following facts are visible on the CT-scans :

- **1** During the initial growth, the tumor is very heterogeneous
- During the first phase of the evolution with the Glivec (cytotoxic drug), the lesion becomes smaller and very homogeneous: this may correspond to a low cellular activity
- 3 Just before the relapse, some heterogeneity appears: a rim of proliferative cells is visible while the center is composed of necrotic cells (darker) Let us note that even if the cellular division has started again, the tumor area has not yet increase.
- When the Glivec failure is obvious (increase of area), clinicians switch treatment. Here, an antiangiogenic treatment is tried: the Sutent.

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- From D) to E), the tumor area decreases again. The tumor darkens: this may correspond to an increase of necrotic cells rate in a tumor. The heterogeneity is also reduced. However, the phenomenon is less important than under Glivec.
- **6** Before the new therapeutic failure, the tumor is very heterogeneous again.
- \Rightarrow RECIST criteria is not a good criteria to evaluate the response to antiangiogenic drug

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EVOLUTION OF THE MASS OF THE TUMOR



FIGURE: Patient A: tumor mass evolution (normalization of integral of grey levels) as a fonction of time in days.

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Main hypothesis I

 Cells are represented by severall populations and we compute the density of the population [6].

No strong biological evidence of this hypothesis.

 \rightsquigarrow Probably, mutated cells do not exist from the beginning but appear during the evolution. The mutation can occur

- before the liver invasion
- just after the colonization when the metastasis is still too small to be visible on CT-scan

■ Angiogenesis plays a crucial role in tumor growth ⇒ we will need to introduce a model to describe it [3]:

- The growth of *P*-cells (proliferatives cells) will be controlled by the quantity of oxygen.
- This oxygen is transported through the bloodstream.
- This bloodstream grows according to a concentration gradient of VEGF (Vascular Endothelial Growth Factor)
- VEGF is itself controlled by the *P*-cells and hypoxia.

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MAIN HYPOTHESIS II

Here, crude model of angiogenesis (to keep the model as simple as posssible) [12]:

 \leadsto We consider direct impact of the $P\mbox{-cells}$ on the speed of migration of oxygen

Spatial expansion of the tumor is governed by a collective velocity induced by the growth of the volume (only a passive movement, no active invasion process is taken into account).
 N.B: This assumption would not be correct for a primary tumor.

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LIST OF VARIABLES

Name	Meaning	Unit
$P_1(t,\mathbf{x})$	Fraction of cells that are both sensitive to imatinib	-
	(Glivec) and sunitinib (Sutent)	
$P_2(t,\mathbf{x})$	Fraction of cells that are resistant to imatinib and	-
	sensitive to sunitinib	
$P_3(t,\mathbf{x})$	Fraction of cells that are both resistant to imatinib	-
	and sunitinib	
$N(t, \mathbf{x})$	Fraction of necrotic cells	-
$S(t, \mathbf{x})$	Fraction of healthy cells	-
$M(t, \mathbf{x})$	Fraction of oxygen // Vascularization	-
$\xi(t)$	Average velocity of oxygen transport in direction	$cm.d^{-1}$
	of the tumor	
$\mathbf{v}(t, \mathbf{x})$	Velocity of the passive movement of the tumor	$cm.d^{-1}$
	under the pressure	
$\Pi(t, \mathbf{x})$	Medium pressure	$kg.cm^{-1}.d^{-2}$

TABLE: List of variables – d = day

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LIST OF PARAMETERS I

			Value	Value
Name	Meaning	Unit	for fit	for fit
			patient A	patient B
γ_0	Tumor cells growth rate	d^{-1}	2.0e-2	6.33e-3
γ_1	Tumor cells apoptosis rate	d^{-1}	8.0e-3	4.46e-2
Cs	Health tissue apoptosis rate	-	10	10
	compared to γ_1			
M _{th}	Hypoxia threshold	-	2	2
μ	Elimination rate of the necrotic	d^{-1}	1.33e-2	8.19e-2
	tissue by the imune system			
ψ	Diffusion rate of the oxygen	$cm^2.d^{-1}$	1.33e-2	3.33e-3
η	Consumption rate of tumor cells	d^{-1}	6.67e-2	8.05e-3
α	Angiogenic excitability	d^{-1}	1.11e-3	8.0e-3
λ	Elimination rate of angiogenic	d^{-1}	2.0e-2	0.68
	growth factor signal			
C_0	Angiogenic capacity of health	d^{-1}	3.33e-2	3.33e-2
	tissue			
k	Tissue permeability	kg ⁻¹ .cm	1	1

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LIST OF PARAMETERS II

			Value	Value
Name	Meaning	Unit	for fit	for fit
			patient A	patient B
δ	Patient sensitivity to imatinib	d^{-1}	7.17e-3	3.45e-3
	and imatinib dosage			
Cg	Sunitinib dose efficiency	-	0.8	0.90
ν	Cytotoxic effects of the suni- tinib	d^{-1}	5.33e-3	3.33e-4
ϵ_{th}	Minimal proportion of tumor cells in a tissue that can be de- tected on scans – Treshold that delimite the tumor area	-	1.0e-2	0.1
Σ_{ini}	$(P_2 + P_3)_{t=0}$	-	3e-06	0.10
q _{ini}	$(P_3/P_2)_{t=0}$	-	7.5e-3	0.41
ξini	Growth factor signal at time <i>t</i> = 0	$cm.d^{-1}$	3.33e-3	0

TABLE: List of parameters – d = day

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GROWTH RATE

R: Numerical parameter – commands the regularisation of the Heaviside step function.

$$\gamma_{pp}(M) = \frac{\gamma_0}{2} \left(1 + \tanh\left(R(M - M_{th})\right) \right)$$
(2.1)

$$\gamma_{pd}(M) = \frac{\gamma_1}{2} \left(1 - \tanh\left(R(M - M_{th})\right) \right)$$
(2.2)

$$\gamma_{sd}(M) = C_S \gamma_1 \max\left(0, -\tanh\left(R(M - M_{th})\right)\right) \quad (2.3)$$



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PDES ON POPULATIONS

$$\partial_t P_1 + \nabla . (\nu P_1) = (\gamma_{pp} - \gamma_{pd}) P_1 - f(t)(1+M) P_1 \qquad (2.4)$$
$$-\nu (1 - g(t))(1+M) P_1$$

$$\partial_t P_2 + \nabla . (\nu P_2) = (\gamma_{pp} - \gamma_{pd}) P_2 - \nu (1 - g(t))(1 + M) P_2 \quad (2.5)$$

$$\partial_t P_3 + \nabla (v P_3) = (\gamma_{pp} - \gamma_{pd}) P_3 \tag{2.6}$$

$$\partial_t N + \nabla . (vN) = \gamma_{pd} (P_1 + P_2 + P_3) + \gamma_{sd} S - \mu (1+M) N \quad (2.7) + f(t)(1+M) P_1$$

- Elimination of necrosis - Action of Glivec f(t)

- Cytotoxic action of Sutent
$$g(t)$$

We assume that :

$$\partial_t S + \nabla . (vS) = -\gamma_{sd} S \tag{2.8}$$

 $+\nu(1-g(t))(1+M)(P_1+P_2)$

By denoting $P = \sum_{i} P_{i}$ and by adding above equations, we obtain:

$$\nabla . \mathbf{v} = \gamma_{\rho\rho} P - \mu (1 + M) N \tag{2.9}$$

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ANGIOGENESIS

$$\partial_t \xi_1 = \alpha \int_{\Omega} \left(1.1 - \frac{\gamma_{pp}}{\gamma_0} \right) (P_1 + P_2) g dx - \lambda \xi_1 \quad (2.10)$$

$$\partial_t \xi_2 = \alpha \int_{\Omega} \left(1.1 - \frac{\gamma_{pp}}{\gamma_0} \right) P_3 dx - \lambda \xi_2 \quad (2.11)$$

By considering the sum, we have :

$$\partial_t \xi = \alpha \int_{\Omega} \left(1.1 - \frac{\gamma_{pp}}{\gamma_0} \right) \left[(P_1 + P_2)g + P_3 \right] \mathrm{d}x - \lambda \xi \qquad (2.12)$$

- $\xi_1(t)$: Average concentration of VEGF.
- $\xi_2(t)$: Average concentration of others growth factors.
- λ : Elimination rate of angiogenic signal.
- α : Production rate of angiogenic signal.

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VASCULARIZATION

$$\partial_t M - \xi \frac{\nabla S}{\|\nabla S\|} \nabla M = \underbrace{C_0 S \left(1 - \frac{M}{2M_s}\right)}_{\text{We impose}} \underbrace{- \widetilde{\eta PM}}_{M} + \psi \Delta M$$

- η : Destruction rate of blood network.
- ψ : Rate of diffusion.

Notice that :

- More the angiogenic signal is important, more the vascularization is transported.
- The vascularization is transported from the health tissue to the tumor.

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System closure

 $\Pi(t,x)$: Pressure v(t,x) : Global speed of cells induced by the pressure. k(t,x) : Tissue permeability Darcy's law :

$$v = -k\nabla\Pi \tag{2.13}$$

where :

$$k(t,x) = k_0 S + k_1 P + k_2 N$$
 (2.14)

Numerically: k = 1A reasonable variation of k not influenced the numericals results.

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Mesh and Method

- 2D cartesian staggered grid $\Omega = [0, L] \times [0, D]$
- Finite volume method
- Fraction of cells are discretized in the center of cells and the velocities are discretized on a middle of each edge of cells



FIGURE: Discretization of unknown variables on typical cell.

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PRESSURE AND VELOCITY CALCULATION

- **1** Initial data : P_1, P_2, P_3, N, S and M at the initial time t = 0
- **2** Δt computating with respect the CFL.
- **B** Pressure computing. By (2.9) and (2.13), we obtain:

$$\begin{cases} -\nabla . (k \nabla \Pi) = F & \text{on } \Omega, \\ \Pi = 0 & \text{on } \partial \Omega, \end{cases}$$
(3.1)

where $F = \gamma_{pp}P - \mu(1 + M)N$. \longrightarrow Classical 5-points scheme.

- Velocity field computing, (thanks to Darcy's law (2.13))
- **5** Cell populations advection.

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ADVECTION EQUATION

Thanks to P + N + S = 1, we can compute S from P and N. Numerically, we compute only the following set of 4 equations:

$$\begin{cases} \partial_t W + \nabla (\mathbf{v}W) = G(W, M) & \text{on } \Omega, \\ W = 0 & \text{on } \partial\Omega, \end{cases}$$
(3.2)

where $W = {}^{t}(P_1, P_2, P_3, N)$, and the function *G* is defined by the relation (2.4) - (2.6) and (2.7). We solve these equations with a splitting time method:

$$\partial_t W + \mathbf{v} \nabla W = 0$$
 then (3.3)
 $\partial_t W = G - WF.$ (3.4)

Non-conservative transport

 \Rightarrow No guarantees that $0 \leq W_i \leq 1$, for $i = 1, \ldots, 4$.

 \Rightarrow High order method (WENO5) in order to minimize this violation.

The equation (3.4) is solved by an Euler or RK3 method.

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VASCULARIZATION COMPUTATION

- ξ(t): Euler or RK3 method to solve the growth factor signal scalar, Eq (2.12).
- **7** M(t, x):(5) is splitted as follows:

$$\partial_{t}M - \psi\Delta M = \xi \frac{\nabla S}{\|\nabla S\|} \nabla M \qquad \text{then} \quad (3.5)$$
$$\partial_{t}M = C_{0}S\left(1 - \frac{M}{2M_{th}}\right) - \eta PM. \qquad (3.6)$$

- Eq. (3.6) is solved by an Euler or RK3 method
- Eq. the equation on vascularization (3.5) is computed as a heat diffusion equation in which the second member is the WENO5 flow.

We impose a Dirichlet condition on the boundary for M:

$$M = 2M_{th} \quad \text{on } \partial\Omega. \tag{3.7}$$

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CFL CONDITION

• CFL condition for WENO5 (including the transport of *M*):

$$\Delta t \leq \min\left(rac{\Delta x}{\max|v_x|}, rac{\Delta y}{\max|v_y|}, rac{\min(\Delta x, \Delta y)}{\max\xi}
ight).$$
 (3.8)

CFL due to the splitting method:

$$\Delta t \leq \min_{i=1,\dots,4} \left(\frac{1}{\max |G_{i,\mathcal{A}} - F|} \right). \tag{3.9}$$

(where $G_{i,A}$ is the linear part of the second member of the advection equations) and (for the vascularization):

$$\Delta t \leq \min\left(\frac{1}{\eta}, \frac{1}{\lambda}\right).$$
 (3.10)

Arbitrary limit:

 $\Delta t \leq 30 \min(\Delta x, \Delta y). \tag{3.11}$

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QUANTITIES OF INTEREST

 $\epsilon_{th}:$ the minimal fraction of tumor cells in a tissue that can be detected on scans.

With this threshold, we can introduce some quantities as tumor area and mass or the sames for each populations cells:

$$\mathcal{A} := \int \mathbb{1}_{\{P+N > \epsilon_{th}\}}(\mathbf{x}) \, \mathrm{d}\mathbf{x}. \tag{3.12}$$

$$\mathcal{A}_J := \int \mathbb{1}_{\{J > \epsilon_{th}\}}(\mathbf{x}) \, \mathrm{d}\mathbf{x}, \quad J \in \{P_1, P_2, P_3, N\}.$$
(3.13)

$$\mathcal{M}_J := \int J \,\mathrm{d}\mathbf{x}, \quad J \in \{P_1, P_2, P_3, N\}, \tag{3.14}$$

$$\mathcal{M} := \int P \, \mathrm{d}\mathbf{x}.\tag{3.15}$$

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CT-SCAN VIEW RECONSTITUTION

Aim: compare the numericals simulations with the CT-scans

 \Rightarrow We need to reconstruct a CT-scan view from the quantities of our model: *P*, *N* and *S*.

 \rightarrow Interpolation of grey level to get a sort of Hounsfield unit (HU) scale.

 \rightarrow Interpolation coefficient are experimentaly fixed

 $(\tau_P=0.65, \tau_N=0.15 \text{ and } \tau_S=0.8)$ thanks to the CT-scan.

We can plotting our numericals results in a grey level by:

$$\tau_P P + \tau_N N + \tau_S S \tag{3.16}$$

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CONSISTENCY OF THE MODEL



FIGURE: Several behaviors that the model is able to reproduce

- ++: A large range of behavior is take into account.
- --: The model is not be predictive.

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PATIENT A

120 points in each direction // CFL: 0.4.



FIGURE: Numerical simulations for patient A: spatial evolution of the lesion with CT-scans reconstitution view.

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PATIENT A



FIGURE: Numerical fit of the tumor area (Patient A)



FIGURE: Population repartition of patient A given by the numerical simulation. The unit of *x*-axis and *y*-axis are *cm* and the grey level represents the fraction of these populations.



FIGURE: Population repartition of patient A given by the numerical simulation. The unit of *x*-axis and *y*-axis are *cm* and the grey level represents the fraction of these populations.



FIGURE: Evolution of the mass (arbitrary unit) and the area (mm^2) of each cellular population and angiogenic signal evolution $(cm.d^{-1})$ given by the numerical simulation.

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PATIENT B I



(A) May 23, 2007 - Day 0

(B) July 25, 2008 - Day 429

(c) Sept 14, 2009 - Day 845

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PATIENT B II



(D) April 06, 2010 – Day 1049

(E) Sept 28, 2010 - Day 1224

(F) May 20, 2011 - Day 1458

FIGURE: Spatial evolution of the patient B metastasis on a series of CT-scans.

As the patient studied in previous section, we try to numerically reproduce the behavior of this tumor with our model.



FIGURE: Patient B: evolution (in FIGURE: Numerical fit of the days) of the tumor area (in mm²) tumor area (on patient B) with a measured on MRIs priori set of parameters.



FIGURE: Numerical simulations a priori for patient B: spatial evolution of the lesion with CT-scans reconstitution view. The units of *x*-axis and *y*-axis are *cm* and the unit of grey scale is arbitrary.

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Adaptation of WENO5 scheme: twin-WENO5



FIGURE: Cut of velocity (in $mm.d^{-1}$) on $y = 6 \ cm$ on 1103^{th} day in function of abscisse x (in cm).

0 0 0 0 0 0 0 \cap 0 Ο 0 Ο Ο Ο Ο 0 0 0 0 0 0 0 ► X

FIGURE: Stencil of the twin-WENO5 scheme.



(A) Spatial aspect of the tumor with CT-scan view reconstitution on day 1366. The units of x-axis and y-axis are *cm* for and the unit of grey scale is arbitrary. (B) Evolution (in days) of tumor area (in mm^2).

FIGURE: Numerical simulation with first correction with twin-WENO5 scheme ($\beta = 0.26$).

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PATIENT B: SIMULATION WITH TWIN-WENO5



FIGURE: Evolution (in days) of the tumor area (in mm^2) given by the model for patient B.

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PATIENT B: SIMULATION WITH TWIN-WENO5



FIGURE: Numerical simulations with twin-WENO5 ($\beta = 0.3$) for patient B: spatial evolution of the lesion with CT-scans reconstitution view. The units of x-axis and y-axis are *cm* and the unit of grey scale is arbitrary.

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CONCLUSION

- Our model is able to reproduce the global behavior of metastasis during the several stage.
- We can mimic both the control of tumor area by the drugs and the relapse.
- Our results are in agreement with the CT-scans.
- We highlight the heterogeneity levels that may be observed in a metastasis. The more the metastases are heterogeneous, the more the relapse is rapidely occurring. This result reinforces the fact that the RECIST criteria is not sufficient to evaluate the efficiency of a treatment.
- Our model is in according with the pharmocological studies on treatment (minimum threshold to have an action, and existence of an optimum dose)



FIGURE: Glivec efficiency on patient A



FIGURE: Glivec efficiency on patient B

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THAT'S ALL

Thanks for your attention.

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